PHACIDIN, A NOVEL Y-PYRONE FUNGAL GROWTH INHIBITOR FROM <u>POTEBNIAMYCES</u> <u>BALSAMICOLA</u> VAR <u>BOYCEI</u> G. A. Poulton^{*} and M. E. Williams

G. A. Poulton^{*} and M. E. Williams Department of Chemistry, University of Victoria Victoria, B. C., V8W 2Y2, Canada.

E. E. McMullan Department of the Environment, Canadian Forestry Service, Pacific Forest Research Centre, Victoria, B. C., Canada.

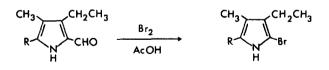
(Received in USA 9 April 1974; received in UK for publication 17 June 1974) Phacidin has been isolated from liquid culture of the canker fungus <u>Potebniamyces</u> <u>balsamicola</u> Smerlis var. <u>boycei</u> Funk, and has been shown to have strong inhibitory effects on the growth of a wide variety of fungi¹. Phacidin is obtained as yellow crystals, mp 112-113°, affords a phenylhydrazone (mp 178°) on treatment with phenylhydrazine hydrochloride, and exhibits a positive Tollens' Test. High resolution mass spectrometry confirms the molecular formula $C_{16}H_{22}O_5$ (found: 294.151; calculated for ${}^{12}C_{16}H_{22}{}^{16}O_5$; 294.147); elemental analysis C: 65.10, H: 7.62% (calculated C: 65.29, H: 7.53%). Phacidin is optically inactive.

The presence of a γ -pyrone nucleus may be inferred from the infra-red absorption at 1690 (C=0), 1620, 1530, 1520 (C=C) and 1235 cm⁻¹ (C-O-C), and from the ultra-violet maxima² at 225 (log ε = 4.34), 279 (3.99) and 320 nm (4.05). The trisubstituted nature of this ring is evident from the spectra: v_{max}^{KBr} 846 cm⁻¹, δ^{CDC13} 7.11 (1H,s); the chemical shift of this proton is assignable to a proton at H₃ or H₅ of the γ -pyrone nucleus (*vide infra*).

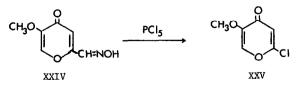
The IR, NMR, and mass spectra reveal that phacidin contains a methoxyl group [1478, 1235, 1041 cm⁻¹; 64.11 (3H,s); m/e 121, 93 ([153-32]⁺ and [121-32]⁺, respectively)], a formyl group [2870, 1730 cm⁻¹; 610.17 (1H,s); m/e 238, 237, 125 ([266-28]⁺, [266-29]⁺, and [153-28]⁺, respectively)], and a third carbonyl group (ν_{max} 1700 cm⁻¹). The nature of this latter substituent was determined by oxidation of phacidin by hydrogen peroxide, neutral KMn0₄, or ruthenium tetroxide to yield n-nonanoic acid, identical in all respects with an authentic sample. The third substituent is thus an n-nonanoyl group: 60.88 (3H,t), 1.1-2.0 (12H,m), and 2.96 (2H,t); m/e 141.128 ([C₈H₁₇C0]⁺, calculated for ${}^{12}C_{9}{}^{1}H_{17}{}^{16}O_{1}$: 141.139) and 153.020 ([M-141]⁺, calculated for ${}^{12}C_{7}{}^{1}H_{5}{}^{16}O_{4}$: 153.019).

2611

The structure XXVI is assigned to phacidin on the basis of the following evidence: i) The methoxyl group can be placed at C_2 on the basis of its lability towards acid³, its characteristic δ value, and its mass spectral behavior; model γ -pyrones derived from maltol and kojic acid, having a C_3 methoxyl substituent, are relatively stable towards acid⁴ and exhibit the OMe resonance in the NMR between & 3.70 and 3.82. All model compounds bearing a C_2 -OMe have been found ⁵ to exhibit $[M-18]^+$ peaks in the mass spectrum; phacidin does not. ii) A carbonyl substituent (CHO or COC_8H_{17}) must be placed at C₆, ortho to the C₅ hydrogen to account for the observed deshielding from the normal resonance position. As can be seen from the accompanying table, protons in the γ -pyrone nucleus are generally observed in the range δ 7.5 - 7.9 when located at positions 2 or 6, while those at positions 3 or 5 are found in the range δ 6.0 - 6.6. Adjacent carbonyl substitution would be expected to deshield such a proton: that this is so is evident from the chemical shift of H₂ in 5-methoxy-4-oxo-4H-pyran-2-carboxaldehyde (XXI). The observed deshielding (0.62 ppm) is comparable to that reported¹³ for substituted furans (0.6 ~ 0.8 ppm). This magnitude of deshielding is not produced by an adjacent alkoxyl group (see compounds XIV, XV), nor is it evident when the carbonyl substituent is not present at an adjacent position (see compound XVI). iii) We have assigned the formyl group to this position (C_2) on the basis of the following evidence: Bromination of phacidin leads to the formation of a bromo-deformyl derivative, as indicated by the loss of the aldehydic proton in the nmr spectrum, and the molecular weight of 345 as indicated by mass spectrometry (molecular ion doublet at 344/346, 8.5% of the base peaks 203/205). This is analogous to the reported 14 deformylation of the pyrrole derivative XXII to yield the 2-bromo-pyrrole XXIII

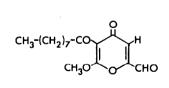


XXII XXIII and is supported by our related replacement of a derivatized C_2 -formyl substituent by halogen under mild conditions¹⁵ (cf XXIV \longrightarrow XXV).



		~	_	TABLE			
CHEMICAL SHIFTS * OF A SERIES OF SUBSTITUTED Y-PYRONES							
<u>pound</u> <u>No</u> .	^R 2	R ₃	R ₅	^R 6	^H 2,6	^H 3,5	Reference
I	Н	н	н	н	7.71	6.35	6
II	CH3	н	н	Ph		6.19(3),6.69(5)) 6
III	СНЗ	н	н	СН3		6.04	6
IV	CH ₃	ОН	н	н	7.73	6.43	6
v	СН3	OCH 3	н	н	7.89	6.23	7
VI	н	CH3	н	н	7.52	6.15	8
VII	сн ₃	OH	н	сн ₃		6.25	9
VIII	OCH2CH3	н	н	СН3		5.95(5)	10
IX	OCH2CH3	CH ₃	H	СНЗ		5.96	10
х	OCH CH 3	Ph	н	сна		6.10	10
XI	OCH2CH3	C1	H	CH ₃		6.10	10
XII	OCH_CH_3	Br	н	CH3		6.10	10
XIII	OCH_CH_	CN	Н	СН3		6.10	10
XIV	OCH	н	н	CH=C(OCH3)Ph		5.5(3),6.8(5)	11
xv	OCH3	н	H	СН3		5.49(3),6.01(5)) 12
XVI	CH3	СООН	н	сн		6.39	This work
XVII	CH2OCH3	н	OCH3	н	7.59	6.45	This work
XVIII	CH_OTHP	н	OCH	н	7.59	6.52	This work
XIX	CH ₂ C1	н	он	н	7.86	6.56	This work
XX	сн ₂ осн ₃	н	он	Ħ	7.85	6.51	This work
XXI	сно	н	OCH3	н	7.69	6.97	This work

The nonanoyl group must then occupy the C_3 position, and phacidin is thus 6-methoxy-5-nonanoyl-4-oxo-4H-pyran-2-carboxaldehyde (XXVI).



XXVI

The authors wish to acknowledge grants in aid of research from the National Research Council of Canada and the University of Victoria, and the kind co-operation of the Pacific Forest Research Centre. This work was carried out during the tenure of an NRC Postgraduate Scholarship. (to M.E.W.)

Footnotes & References:

- 1. A. Funk and E. E. McMullan, Can. J. Microbiol., in press.
- 2. H.-D. Becker, Acta Chem. Scand. 16, 78 (1962) reports λ_{max} 216-218 (log ε = 4.1) and 266-272 nm (3.9) for similar γ -pyrones.
- Full experimental details for this and other chemical transformations will be reported in a subsequent full paper.
- 4. See for example the work quoted in ref. 2.
- 5. D. McGillivray, G. A. Poulton, and M. E. Williams, unpublished results; presented at the 57th Canadian Chemical Conference, June 5, 1974, Regina, Sask.
- 6. C. T. Mathis, and J. H. Goldstein, Spectrochim. Acta , 20, 871 (1964).
- V. F. Bystrov, V. P. Lezina, V. M. Dashunin, and M. S. Tovbina, <u>J. Gen. Chem. USSR</u>., 34, 2918 (1964).
- 8. D. W. Mayo, P. J. Sapienza, R. C. Lord and W. D. Phillips, J. Org. Chem., 29, 2682 (1964).
- 9. K. Sato, S. Inoue, and M. Ohashi, Bull. Chem. Soc. Japan, 46, 1288 (1973).
- 10. T. Kato, Y. Yamamoto, and S. Takeda, <u>Chem. Pharm. Bull.</u>, <u>21</u>, 1047 (1973).
- 11. M. P. Wachter and T. M. Harris, Tetrahedron, 26, 1685 (1970).
- 12. P. Beak and H. Abelson, <u>J. Org. Chem</u>., <u>27</u>, 3715 (1962).
- 13. L. M. Jackman and S. Stornhell, "Applications of Nuclear Magnetic Resonance Spectroscopy in Organic Chemistry", Second Edition, Pergamon Press, Oxford, 1969, p. 214.
- 14. A. Markovac and S. F. MacDonald, Can. J. Chem., 43, 3364, (1965).
- 15. G. A. Poulton and M. E. Williams, Can. J. Chem., submitted for publication.